Iron is essential for many biological processes, including oxygen delivery, and its supply is tightly regulated. Hepcidin, a small peptide synthesized in the liver, is a key regulator of iron absorption and homeostasis in mammals. Hepcidin production is increased by iron overload and decreased by anemia and hypoxia; but the molecular mechanisms that govern the hepcidin response to these stimuli are not known. Here we establish that the von Hippel–Lindau/hypoxia-inducible transcription factor (VHL/HIF) pathway is an essential link between iron homeostasis and hepcidin regulation in vivo. Through coordinate downregulation of hepcidin and upregulation of erythropoietin and ferroportin, the VHL-HIF pathway mobilizes iron to support erythrocyte production.

Introduction
Iron metabolism and homeostasis are dependent on the proper regulation of hepcidin, a small antimicrobial peptide expressed in the liver that has only recently been cloned and characterized (1). Genetic studies in mice lacking or overexpressing hepcidin suggest that this peptide inhibits both iron absorption in the small intestine and the release of recycled iron from macrophages (2, 3). To limit iron export, hepcidin blocks ferroportin, the sole known iron exporter, by binding to it and causing its internalization and degradation (1). Hepcidin upregulation in response to chronic inflammatory stimuli (e.g., cancer, infection, autoimmunity) leads to the anemia of chronic disease (4–7), while hepcidin downregulation is associated with the iron overload disorder of hemochromatosis (8–11).

Given the links among oxygen transport, erythropoiesis, and iron metabolism, there are potential associations between the physiology of hypoxic response and the control of iron availability. To gain insight into potential hypoxia-dependent molecular mechanisms that might control hepcidin expression in iron homeostasis, we examined transcriptional control of the hypoxic response in the liver by the hypoxia-inducible transcription factors (HIFs), key elements in the response to hypoxia (12). We hypothesized that this pathway could play a role in hepcidin regulation because: (a) iron deprivation and hypoferic anemia lead to poor tissue oxygenation; and (b) HIF levels are partially regulated by the tumor suppressor protein von Hippel–Lindau (VHL). The VHL protein regulates turnover of components of the HIF transcription factors through a process that requires molecular iron.

HIF is a heterodimer whose expression is regulated posttranslationally. Three regulatory HIF subunits have been cloned and named HIF-1α, HIF-2α (13–15), and HIF-3α (16). In the presence of oxygen, the regulatory subunits are modified by iron-dependent prolyl hydroxylases (PHDs) and are then degraded through the ubiquitin/proteasome pathway via its interaction with VHL. Under hypoxia or following iron chelation, PHD activity is inhibited. The regulatory subunit then accumulates and translocates into the nucleus, where it binds to aryl hydrocarbon receptor nuclear translocator/HIF-1α (ARNT/HIF-1α), which is constitutively expressed. The heterodimeric HIF binds to the hypoxia-response elements (HREs) of target gene regulatory sequences, resulting in the transcription of genes implicated in the control of metabolism and angiogenesis, as well as apoptosis and cellular stress (12). Erythropoietin (EPO) is one of these direct target genes.

In this study, we establish HIF as an essential link between iron fluctuation and hepcidin regulation. The fact that hepcidin, ferroportin, and EPO under the control of VHL/HIF pathway allows a coordinated response for controlling iron metabolism and oxygen transport, regulating the processes that provide iron for physiologic processes such as erythropoiesis. These data provide what we believe to be the first molecular evidence for a coordination between the process of iron uptake and hypoxic response.

Results
HIF-1α is stabilized in the liver of iron-starved mice. Mutant mice with iron-deficiency anemia or rats fed an iron-deficient diet exhibit lower hepatic hepcidin expression than control animals (6, 17). Studies in patients with iron-deficiency anemia have also demonstrated significantly reduced hepcidin levels relative to those of healthy subjects (18).

We observed that iron starvation of mice of the inbred C57BL/6 strain resulted in a strong decrease in hepatic hepcidin mRNA expression compared with that in control mice fed a normal diet (Figure 1A). Mice in this same study had a clear increase in HIF-1α levels in the liver in response to the iron deficiency (Figure 1B). This demonstrates that HIF-1α expression in the liver is influenced by dietary iron depletion.

Decreased regulation of hepcidin by nutritional iron when HIF-1α is deleted in hepatocytes. HIF-1α was specifically inactivated in hepatocytes by crossing C57BL/6 backcrossed HIF-1αfl×/− mice...
with Albumin-Cre C57BL/6 transgenic mice. Deletion efficiency in total liver was approximately 70% as determined by quantitative PCR (data not shown). This indicates a very high rate of hepatocyte deletion, since it is estimated that other cell types, such as endothelial cells and Kupffer cells, make up approximately 30% of the cells in the liver (19). Under a standard iron-rich laboratory diet, HIF-1α expression was not detectable in the liver (Figure 1B); and no significant differences in hepatic hepcidin expression were observed between WT and Albumin-Cre/HIF-1α\(^{flox/flox}\) mice (Figure 1C).

After 20 days of feeding with an iron-deficient diet, Albumin-Cre/HIF-1α\(^{flox/flox}\) mice expressed 10-fold more hepcidin than WT littermates (Figure 1C), indicating that deletion of HIF-1α affects the physiological mechanisms governing hepcidin downregulation during nutritional iron deficiency. However, we found that elimination of HIF-1α alone is insufficient to fully compensate for the hepcidin reduction caused by an iron-deficient diet (Figure 1C), suggesting that other factors also contribute to the observed hepcidin downregulation.

Deletion of VHL in hepatocytes results in polycythemia and iron deficiency. The VHL pathway that controls HIF-1α levels also regulates HIF-2α, which in turn is thought to have unique roles in regulating liver tumorigenesis and EPO expression (20). The VHL tumor suppressor regulates the oxygen-sensitive turnover of both HIF-1α and HIF-2α transcription factors. To investigate the effect of altering the stability of both factors in vivo, we deleted VHL in the liver by crossing C57BL/6 VHL\(^{flox/flox}\) with Albumin-Cre C57BL/6 transgenic mice.

Growth deficiency, polycythemia, and reduced survival occur when VHL is deleted in the liver in a mixed inbred strain genetic background (BALB/c, 129SvJ, and C57BL/6) (21). In the pure C57BL/6 background, mice homozygous for the VHL mutation (Albumin-Cre/VHL\(^{flox/flox}\)) in the liver demonstrated alopecia and grew to only half the weight of their WT littermates (Figure 2A), dying between 5 and 7 weeks of age with severe hepatomegaly and splenomegaly (Figure 2B). VHL deletion efficiency in total liver was approximately 50% as determined by quantitative PCR evaluation of genomic DNA (data not shown).

The livers of Albumin-Cre/VHL\(^{flox/flox}\) mice exhibited steatosis (pathologic fat accumulation), with inflammatory cell infiltration as well as foci of necrosis due to fibrin clots (Figure 2B); however, no fibrotic changes were noted. Albumin levels indicative of normal hepatic synthetic function were observed (data not shown). Serum levels of the HIF target gene EPO were increased, reflecting the strong upregulation of EPO mRNA in the liver of Albumin-Cre/VHL\(^{flox/flox}\) compared with WT mice (Figure 2C).

Increased EPO levels are consistent with the increased erythrocytosis and elevated hematocrit observed in the Albumin-Cre/VHL\(^{flox/flox}\) mice (Figure 2C). Although total hemoglobin levels were elevated (Figure 2C), mean cell hemoglobin (MCH) was decreased in Albumin-Cre/VHL\(^{flox/flox}\) mice compared with WT mice (Figure 2D). This result indicates a relative iron deficiency in the animals. The observation was corroborated by the hypochromia, poikilocytosis, and microcytosis observed in blood smears from the Albumin-Cre/VHL\(^{flox/flox}\) mice (Figure 2D).

Total iron (Figure 2E) and ferritin (Figure 2F) levels in the livers of Albumin-Cre/VHL\(^{flox/flox}\) mice were also greatly reduced relative to those of WT mice. The spleen, another reservoir of iron, was also strongly iron deficient according to Perls Prussian blue staining, as shown in Figure 2G.

The phenotype of the Albumin-Cre/VHL\(^{flox/flox}\) mice is similar in some regards to that of humans with Chuvash polycythemia, in which a homozygous mutation (598C→T) in the VHL gene leads to elevated normoxic levels of HIF-1, serum EPO, and hemoglobin and premature mortality related to cerebral vascular events and peripheral thrombosis (22, 23).

HIF-1 binds to the promoter of hepcidin in vivo and reduces its expression in the liver. We detected 2 candidate consensus HREs ([A/G]CGTG) in the murine hepcidin promoter (24) (Figure 3A). We performed a chromatin IP (ChIP) assay on liver tissues extracted from WT and Albumin-Cre/VHL\(^{flox/flox}\) mice. Primers flanking HIF-1α consensus binding sites specifically amplified DNA sequences immunoprecipitated by an HIF-1α antibody, indicating that HIF-1α is able to bind the murine hepcidin promoter in vivo (Figure 3A), which supports the assumption of direct regulation of hepcidin by HIF-1.

Examining the human hepcidin promoter sequence, we detected 3 candidate consensus HREs. We performed a ChIP on human HEK293 cells stimulated with the iron chelator desferrioxamine mesylate (DFO) in order to stabilize HIF-1α. In these conditions, primers flanking HIF-1α consensus binding sites specifically amplified DNA sequences immunoprecipitated by HIF-1α antibody, showing that HIF-1α is able to bind the hepcidin promoter in human cells as well as mouse tissues (Figure 3B). We next examined whether the putative classical and conserved HRE (first of 2 HREs arranged as inverted repeats) in the human hepcidin promoter was important for the iron-dependent regulation of hepcidin expression. We cloned the promoter region of the human hepcidin encompassing the HREs into a luciferase reporter construct. The HIF agonist DFO strongly inhibited luciferase activity of this plasmid (pGL3-Hepc/HRE) (Figure 3C), suggesting that HIF binding to the hepcidin promoter suppresses hepcidin gene transcription. To verify that the putative Hepc/HRE located at −582 bp is the target of repression by HIF, we intro...
duced a point mutation (5'-CAA-TG-3' instead of 5'-CACGTG-3') into this sequence (Figure 3D). The activity of the corresponding plasmid that bears the mutation (pGL3-Hepc/muHRE) was increased compared with that of the pGL3-Hepc/HRE plasmid. These results demonstrate that the sequence covering the consensus HIF-1 binding site is involved in the downregulation of hepcidin gene transcription by HIF.

We next assessed whether the increased amounts of HIF-1α in the liver, and its binding to the hepcidin promoter regulate the hepcidin gene in murine tissues. As shown in Figure 3E, Hepc1 mRNA levels measured in the liver of Albumin-Cre/VHL<sup>fl/fl</sup> mice were markedly decreased compared with those of WT littermates. Analysis of protein levels confirmed that levels of the hepcidin peptide were also lower in Albumin-Cre/VHL<sup>fl/fl</sup> mice (Figure 3E).

Downregulation of Hepc1 mRNA in Albumin-Cre/VHL<sup>fl/fl</sup> mice was eliminated, i.e., transcript levels returned to normal values, in Albumin-Cre/VHL<sup>fl/fl</sup>/ARNT<sup>fl/fl</sup> double mutant (double-knockout) mice (Figure 3F). These mice have a hepatic deletion of both the VHL gene, whose absence causes increased HIF activity, and the ARNT gene, which is required for dimerization and thus activation of HIF-mediated transcription. The restoration of hepcidin to normal levels in the livers of these double...
The research article published in The Journal of Clinical Investigation on July 2007 discusses the role of hepcidin in regulating iron metabolism. It highlights that hepcidin downregulation in Albumin-Cre/VHL-flx/flx mice is specifically due to the stabilization of HIF transcription factors (i.e., HIF-1α and HIF-2α) and not to other functions of the VHL gene.

As Albumin-Cre/VHL-flx/flx mice exhibit hepatic inflammation and steatosis, the article examines the levels of the proinflammatory cytokine IL-6, which is known to stimulate hepcidin expression and may contribute to the anemia of chronic inflammation. Significantly elevated IL-6 and IL-1β levels were found in the Albumin-Cre/VHL-flx/flx mice (Figure 3G), although this change did not prevent the decreased hepcidin expression described above. These findings indicate that IL-6- or IL-1- mediated stimulation of hepcidin expression is subordinate to the suppression caused by increased HIF transcriptional activity.

Upregulation of ferroportin in Albumin-Cre/VHL-flx/flx mice. It has recently been shown ex vivo that hepcidin binds ferroportin, the only known iron exporter, thereby trapping iron within enterocytes and macrophages. Through this mechanism, increased ferroportin expression is observed in brush border enterocytes of the duodenum of Albumin-Cre/VHL-flx/flx mice (Figure 4A). Similarly, increased ferroportin expression is seen in Kupffer cells of the liver, and this was also observed in the hepatocytes (Figure 4A).

Western blot analysis confirmed the increase in ferroportin expression in whole liver extracts from Albumin-Cre/VHL-flx/flx mice relative to WT mice (Figure 4B). Increased ferroportin levels in the hepatocytes may in part reflect a small but statistically significant increase in hepatic ferroportin mRNA expression detected in Albumin-Cre/VHL-flx/flx mice (Figure 4C). While ferroportin expression in the duodenum could result from the decrease in systemic hepcidin expression or from the systemic iron deficiency itself,

Figure 3
Binding of HIF-1 to the promoter of hepcidin and downregulation of hepcidin in Albumin-Cre/VHL-flx/flx mice. (A) Sequence of murine (C57BL/6) hepcidin promoter; HREs are in bold; arrows indicate primers selected for ChIP. (B) DFO (150 μM) induces binding of HIF-1 as shown by ChIP assay. (C) Luciferase-reporter constructs under the control of the regulatory region of the human hepcidin gene. HEK293 cells transiently transfected with pGL3 basic or pGL3-Hepc/HRE vector. (D) The “native” (CCACGTG) and mutated (CAA-TG) HREs (indicated by an X) are shown. HEK293 cells were transiently transfected with pGL3 basic, pGL3-Hepc/HRE, or pGL3-Hepc/mutHRE. (E) Hepsin mRNA expression in livers of WT and Albumin-Cre/VHL-flx/flx by real-time RT-PCR (n = 8). HIF-1 and hepcidin expression in liver extracts of WT and Albumin-Cre/VHL-flx/flx mice. (F) Hepsin mRNA expression in livers of WT, Albumin-Cre/VHL-flx/flx, and Albumin-Cre/VHL-flx/flx/ARNT-flx/flx (VHL–/–ARNT–/–) mice (n = 4). (G) IL-6 and IL-1β mRNA levels in livers of WT and Albumin-Cre/VHL-flx/flx mice.
expression of ferroportin in the hepatocyte may reflect a direct tissue-autonomous effect of VHL-deficient cells. Indeed, as previously reported, iron export in hepatocytes is likely independent of the posttranslational regulation of ferroportin by hepcidin (26, 27).

Discussion
Hepcidin is suppressed by both anemia and hypoxia (7). Cellular oxygen sensing and hypoxia-induced transcription are largely mediated by the HIFs. The stabilization of HIF-1 by iron chelators has been well established in vitro. However, our demonstration of the in vivo induction of HIF-1 by iron deficiency, and the associated downregulation of hepcidin when HIF levels are elevated, suggests that HIF may be one of the missing links between iron homeostasis and hepcidin regulation. The ability of HIF-1α to bind to and negatively transactivate the hepcidin promoter suggests a direct repressor effect. HIF-1α has already been reported to repress the transcriptional activity of genes such as alpha-fetoprotein (AFP) (28) and carbamoyl-phosphate synthetase 2, aspartate transcarbamylase, and dihydroorotatase (CAD) (29).

Our data show that elimination of HIF-1 alone in adult mice is insufficient to fully compensate for the hepcidin reduction induced by iron depletion. The striking downregulation of hepcidin observed in Albumin-Cre/VHL^{flax/flax} mice, in which both HIF-1α and VHL were achieved by cross-breeding Albumin-Cre transgenic mice (The Jackson Laboratory) with VHL^{flax/flax} mice (backcrossed in a pure C57BL/6 genetic background). In all experiments, littermates from the same breeding pair were used as controls. Albumin-Cre/VHL^{flax/flax} and Albumin-Cre/VHL^{flax/flax}/ARNT^{flax/flax} mice used in the experiments shown in Figure 3C are in a mixed BALB/c, 129SvJ, and C57BL/6 genetic background. Mice were maintained on a standard rodent diet (formula LM-458, 7912; Harlan Teklad) containing 284 mg iron/kg. In certain experiments, mice were fed a low-iron (3–6 mg/kg food) diet (Formula TD 80396; Harlan Teklad) for 3 weeks.

Hematological analysis. Blood was obtained by retro-orbital bleeding and collected in EDTA-K2 tubes (Capiject T-MLH; TERUMO). Blood cell counts and erythrocyte parameters were determined using a MAXM Coulter (Beckman Coulter) automatic analyzer.

Immunohistochemistry. Tissues were fixed in 10% formalin sectioned in paraffin, then subjected to microwave antigen retrieval and immunohistochemistry (Vector Blue Alkaline Phosphatase Kit; Vector Laboratories) with primary antibodies against ferroportin (Alpha Diagnostic International) at 1:200. Iron detection was performed with Perl's Prussian blue and nuclear red counter stain.
Iron quantification. Tissue iron was quantified colorimetrically by a modification of the method of Torrance et al. (33). Fifty- to 100-mg liver extracts were digested overnight in 100 μl 3 M HCl/10% trichloroacetic acid at 65 °C, followed by the addition of 200 μl chromogen (0.01% bathophenanthroline sulfonate, 0.1% thiglycolic acid, and 4 M sodium acetate). For a standard curve, serial dilutions of a ferric iron standard (Sigma-Aldrich) were used. Color was allowed to develop for 15 minutes and measured at 535 nm.

RT and real-time quantitative PCR. First-strand synthesis was obtained from 1 μg of total RNA isolated with TRI reagent (Molecular Research Center Inc.) by the SuperScript system (Invitrogen) employing random primers. For real-time PCR analyses, cDNAs were diluted to a final concentration of 10 ng/μl and amplifed in a SYBR Green TaqMan Universal Master Mix (Applied Biosystems). Fifty nanograms cDNA was used as template to determine the amount of mRNA by real-time PCR in triplicate (ABI Prism 7700 sequence detection system; Applied Biosystems), using specific primers with the following sequences: hepcidin forward, 5′-TGTCCTCCTCTCTCTCTCCT-3′; hepcidin reverse, 5′-CTCCGTAGCTGCTCCTCATCTGTTG-3′; ferroportin forward, 5′-CTACATGAGGATGTTGACCACTGA-3′; ferroportin reverse, 5′-ACTGGAGAAGCCTATGTCAATCTG-3′; EPO forward, 5′-AATGGAGGTGGAAGAACCAGGCAT-3′; EPO reverse, 5′-GGGACGCTGGAAGTTGACCTACGTA-3′; β-actin forward, 5′-AGGCCAGAAGCACAGGAGG-3′; β-actin reverse, 5′-TACATGGCTGGGTTTGAAG-3′.

ChIP assay. For the in vivo ChIP assay, approximately 130 mg of livers from WT and VHL−/− mice were extracted and were submitted to the protocol developed by Farnham’s laboratory and described at http://genecenter.ucdavis.edu/farnham/farnham/protocols/tissues.html.

Primers flanking the putative HREs within the murine Black6 Hpe promoter were: −2366-mHepC-F, 5′-CCATCATTCTTAACTGGGCTC-3′, and −1915-mHepC-R, 5′-GCCATTGTACACGTACATCA-3′.

Western blot studies. Livers were harvested, ground in a mortar, and washed with PBS and proteins extracted with RIPA buffer. Twenty micrograms of total (for ferritin and ferroportin) or nuclear extracts (for hepcidin and HIF) were loaded on a 10% (ferritin and ferroportin), 12% (hepcidin), or 7% (HIF) gels, and Western blot analysis was performed using standard methodologies. Primary antibodies were rabbit anti-mouse Hcpdin (Alpha Diagnostic International), rabbit anti–HIF-1α (Novus Biologicals), rabbit anti-ferroportin (Alpha Diagnostic International), and rabbit anti–ferritin heavy chain (Abcam) at a concentration of 1:1,000.

Transient transfection and luciferase assay. The hepcidin promoter luciferase reporter plasmid (pGL3-HRE) was constructed by cloning a PCR fragment of the human hepcidin upstream region (−762 bp to −20 bp) into a pGL3 vector (Promega) by conventional cloning procedures to yield pGL3-Hepc/HRE. A mutation of the putative HRE, in which the CG in the CACGTTG motif was replaced by an A, was generated by site-directed mutagenesis using the QuikChange Site-Directed Mutagenesis kit (Stratagene) as indicated by the manufacturer. The point mutation in the corresponding plasmid pGL3-Hepc/mutHRE was verified by sequencing. HEK293 cells were transfected by Lipofectamine 2000 (Invitrogen) with 2 μg of pGL3, pGL3-Hepc/HRE, or pGL3-Hepc/mutHRE, according to the manufacturer’s protocol. The following day, cells were incubated with DFO at 150 μM for 18 hours. Cells were then washed with PBS and lysed with the reporter gene assay lysis buffer (Roche), and luciferase assay was performed using the Bright-Glo Luciferase Assay kit (Promega).

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